



EFFECT OF *SMILAX CHINA* LINN ON TESTICULAR ANTIOXIDANT ACTIVITY AND SPERMATOLOGICAL PARAMETERS IN RATS SUBJECTED TO FORCED SWIMMING STRESS

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Article Received on: 06/09/12 Revised on: 01/10/12 Approved for publication: 06/11/12

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ABSTRACT

The aim of the present study was to investigate the potential benefits of ethanolic extract of *Smilax china* Linn. on testicular antioxidant activity and spermatological parameters in rats subjected to forced swimming stress. Animals of the experimental groups except vehicle control were subjected to forced swimming stress (FSS) 15 min/day for 52 days. Animals were pretreated with two doses of ethanolic extract of *Smilax china* rhizomes (100 and 200 mg/kg b.w., p.o.) for 15 days prior to the starting of FSS and were continued further for 52 days along with induction of stress. Testicular SOD, catalase and lipid peroxidation were determined. The cauda epididymis was isolated and sperms were released into saline and sperm count, viability, morphology and motility were analysed. Rats with forced swimming stress showed a significant increase in lipid peroxidation and decrease in testicular SOD, catalase, sperm count, viability and motility. Many abnormal forms of sperms were seen. Rats pretreated with ethanolic extract of *Smilax china* rhizomes at both doses significantly prevented the stress induced changes. Hence, ethanolic extract of *Smilax china* rhizomes at both doses showed good protection against testicular antioxidant activity and spermatological parameters in rats subjected to forced swimming stress.

Keywords: *Smilax china* rhizomes, Forced swimming, Testicular antioxidant, Spermatological parameters

INTRODUCTION

Oxidative stress participates in the autogenesis of more than 100 diseases. Reactive oxygen species formed in the oxidative stress react with biomolecules such as lipids, nucleic acids and proteins, changing their structure and thus their function, leading to cell damage and alters physiological functions^{1,2}. Approximately 15% of couples attempting to conceive are clinically infertile and male infertility contributes to 10 – 30% of those cases³. Common causes include excessive alcohol consumption, drugs, toxicant exposure, restraint, excessive exercise and conditions like varicocele, cryptorchidism, testicular torsion, or endocrinopathy. All of which results in testicular damage, impairment of spermatogenesis and sperm function^{4,5}. Oxidative stress is also a major factor contributing to male infertility.

Smilax china Linn. (Family: Liliaceae) also known as chopchini, madhusnuhi is a popular herb in Ayurveda⁶. It is imported from China and Japan. In traditional medicine the plant is used to prevent and treat a wide variety of diseases such as diuretic, liver disorders, skin diseases, seminal weakness, aphrodisiac, venereal diseases, infertility etc⁷. Experimental reports indicated that *Smilax china* possessed anti inflammatory⁸, anti nociceptive⁸, anticancer⁹, immunomodulatory¹⁰, anti microbial activities¹¹. The various pharmacological activities were due to the presence of chemical constituents like saponins, alkaloids, flavonoids etc. The present study was aimed to evaluate the potential benefits of ethanolic extract of *Smilax china* on testicular antioxidant and spermatological parameters in rats subjected to forced swimming stress.

MATERIALS AND METHODS

Plant material

The rhizomes of *Smilax china* Linn were authenticated and the ethanolic extract was obtained from Green chem herbal extracts and formulations, Bangalore, Karnataka, India (Batch no: SCR/RD/04)

Animals

Experimental study was carried out using adult male Wistar albino rats weighing between 170–200 g. The animals were procured from Drug Testing Laboratory, Bangalore. The animals were housed in polypropylene cages and were maintained clean and hygiene. Animals were acclimatized in a room 12–12 h dark–light cycle, maintained at temperature 25±2°C and humidity 50±5%. The rats were fed with commercial pelleted rat feed (Gold Mohur Lipton India Ltd.) and water *ad libitum*. The animal caring and handling were done according to the CPCSEA guidelines. The Institutional Animal Ethics Committee (IAEC/NCP/17/10) approved the above study at Nargund College of Pharmacy, Bangalore.

Acute oral toxicity study

The acute oral toxicity study was performed according to the OPPTS (Office of Prevention, Pesticides and Toxic Substance) guidelines following the Up and down procedure¹².

Forced swimming stress

The rats were placed individually in an acrylic plastic pool (90 cm×45 cm×45 cm) filled with water up to a depth of 37cm and was made to forced swim for 15 min/day for 52 days at ambient room temperature^{13,14}. Animals were divided into four groups, six animals in each group.

Group I- Vehicle control - distilled water, orally (5 mL/kg body weight)

Group II- Forced swimming stress (15 min/day) for 52 days.

Group III and IV –Rats were pretreated with oral administration of ethanolic extract of *Smilax china* (EESC) rhizomes (100 and 200 mg/kg b.w., p.o.) respectively for 15 days prior to the starting of forced swimming stress and was continued further for 52 days along with induction of stress. From 16th day the animals were subjected to forced swimming stress for 15min/day for 52 days half an hour after administration of the extract.

After the last stress session, control and other group of animals were sacrificed by cervical dislocation. Testis were removed, weighed and placed in KCl (10% w/v) solution, homogenated using a homogenizer and centrifuged at 4000

rpm for 10 min. Liver homogenate was analysed for lipid peroxidation. The supernatant were separated and estimated for SOD and catalase. The cauda epididymides were isolated and spermatological parameters such as sperm motility, viability, count and morphology were performed^{15,16}.

Statistical Analysis

Data are expressed as mean \pm SEM and were analysed statistically using one way ANOVA followed by Dunnett's *t* test. Results were considered significant when P values were <0.05 .

Table 1 Effect of ethanolic extract of *Smilax china* (EESC) rhizomes on testicular antioxidant enzymes (SOD and catalase units/mg of protein) and lipid peroxidation (n mol of MDA/mg of protein) in rats

| Groups | SOD | Catalase | Lipid peroxidation |
|----------------------------|----------------------------------|---------------------------------|----------------------------------|
| Vehicle control | 152.9 \pm 7.73 | 1.11 \pm 0.12 | 102.6 \pm 1.18 |
| FSS | 94.37 \pm 0.98*** ^a | 0.67 \pm 0.01*** ^a | 152.1 \pm 0.68*** ^a |
| FSS + EESC (100 mg/kg b.w) | 127.0 \pm 0.67*** ^b | 0.87 \pm 0.01*** ^b | 122.4 \pm 0.15*** ^b |
| FSS + EESC (200mg/kg b.w) | 142.8 \pm 0.76*** ^b | 1.05 \pm 0.03*** ^b | 113.2 \pm 0.96*** ^b |

Values are expressed as Mean \pm SEM. Data were analyzed by one way ANOVA followed by Dunnett's *t* test. n = 6.

^a Comparison made with vehicle control group. ^b Comparison made with FSS group. *** P<0.001

Table 2 Effect of ethanolic extract of *Smilax China* rhizomes on spermatological parameters in rats

| Groups | Epididymis sperm count ($\times 10^6$ /mL) | Sperm viability (% of viable sperms) | Sperm motility (% of rapidly progressive motile sperms) | Sperm morphology |
|----------------------------|---|--------------------------------------|---|---------------------|
| Vehicle control | 107.0 \pm 0.36 | 45.0 \pm 0.81 | 24.0 \pm 0.68 | Normal |
| FSS | 77.95 \pm 0.52*** ^a | 13.0 \pm 0.57*** ^a | 3.45 \pm 0.12*** ^a | More abnormal forms |
| FSS + EESC (100 mg/kg b.w) | 94.10 \pm 0.25*** ^b | 21.33 \pm 0.61*** ^b | 16.26 \pm 0.18*** ^b | Few abnormal forms |
| FSS + EESC (200 mg/kg b.w) | 98.14 \pm 0.25*** ^b | 39.0 \pm 0.36*** ^b | 23.50 \pm 0.42*** ^b | Normal |

Values are expressed as Mean \pm SEM. Data were analyzed by one way ANOVA followed by Dunnett's *t* test. n = 6.

^a Comparison made with vehicle control group. ^b Comparison made with FSS group. *** P<0.001

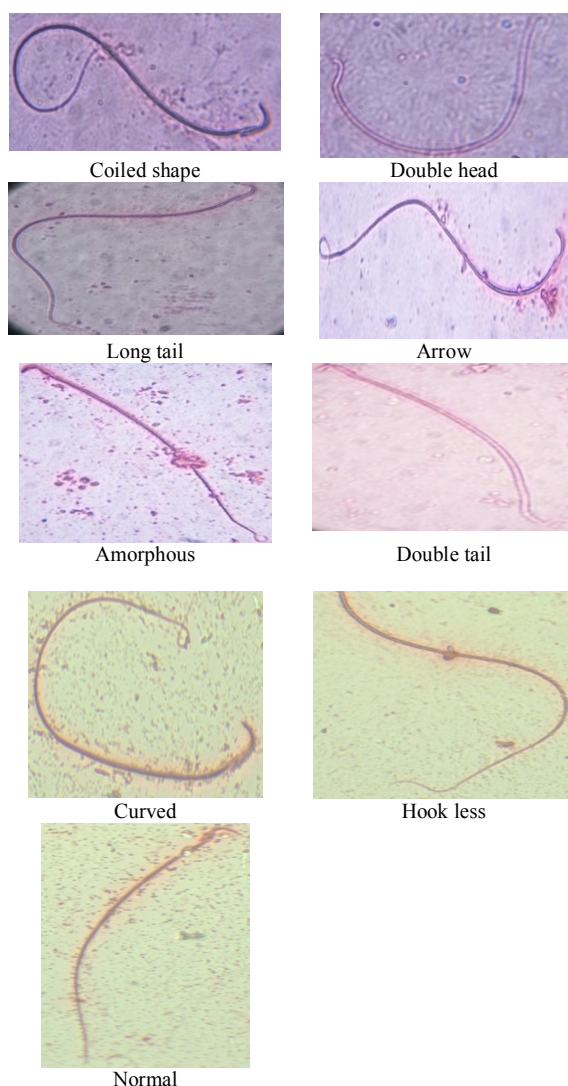


Figure 1 Abnormal forms of sperm

RESULTS

Acute oral toxicity study

EESC rhizomes showed no mortality in rat's upto a dose of 5000 mg/kg b.w. Hence 1/50th and 1/25th of the dose i.e. 100 and 200 mg/kg b.w was selected for improving antioxidant and spermatological parameters in forced swim stressed rats.

Forced swimming stress

Forced swimming stressed rats showed a significant decrease ($P<0.001$) in the SOD, catalase and increase in lipid peroxidation levels when compared with vehicle control group. Prior and continued treatment with EESC rhizomes at both doses of 100 and 200 mg/kg b.w prevented the fall of testicular antioxidant enzymes SOD and catalase levels ($P<0.001$). The lipid peroxidation damage was also reduced ($P<0.001$) against stress induced changes (Table 1)

A significant decrease ($P<0.001$) in the epididymis sperm count, sperm viability and sperm motility was seen in stressed rats. More number of abnormal forms such as banana, curved, double tail, and hookless, amorphous and double head were also seen. Prior and continued treatment with EESC rhizomes at doses 100 and 200 mg/kg b.w showed a significant improvement ($P<0.001$) in the epididymis sperm count, sperm viability and sperm motility when compared with forced swimming stress group (Table 2). Very few abnormal forms were seen at 100 mg/kg and at 200 mg/kg b.w almost normal forms were seen.

DISCUSSION

Forced swimming, a good physical exercise model, is also considered as a physical stressor. During physical exercise, oxygen utilization increases 10-15 folds and it is well established that reactive oxygen species (ROS) generation is a direct function of the rate of oxygen utilization and oxygen reperfusion is another process of ROS imposition. From review of literature, it has been indicated that remarkable dysfunctions are noted in male reproductive system, inhibiting spermatogenesis due to intensive exercise¹⁴. Ethanolic extract of *Smilax china* rhizomes was evaluated for

improving male fertility parameters in stressed rats using forced swimming stress model.

SOD is the first antioxidant enzyme to deal with oxyradicals by accelerating the dismutation of superoxide (O_2^-) to hydrogen peroxide. CAT is a peroxisomal haem protein that catalyses the removal of H_2O_2 formed during the reaction catalysed by SOD. Thus, SOD and CAT acts mutually supportive antioxidative enzymes, which provide protective defense against reactive oxygen species. These ROS are very unstable and highly reactive. They become stable by acquiring electron from nucleic acids, proteins, carbohydrates and lipids, there by a cascade of chain reaction are initiated resulting in cellular damage and causes lipid peroxidation¹⁷. Testes of forced swimming stressed rats showed a significant decrease in the SOD and catalase enzyme levels may be due to excessive generation of free radicals or low levels of testosterone since testosterone promotes the synthesis of antioxidant enzymes in sex organs^{18,19}. EESC rhizomes at both doses showed significant protection in antioxidant enzyme levels of SOD and catalase and showed normal basal levels.

Malondialdehyde a secondary product of lipid peroxidation is a major reactive aldehyde; higher levels leads to peroxidation of biological membranes²⁰ or due to high rate of catecholamine secretion that generate free radicals either through auto oxidation or through metal ion or superoxide-catalyzed oxidation¹⁴. The present data revealed significant increase in lipid peroxidation level which may be due to oxidative stress resulting in cellular damage. Previous studies also have shown that intensive exercise is linked with testicular oxidative stress²¹. The EESC extracts at both doses were able to prevent the lipid peroxidation due to free radical scavenging properties.

Spermatogenesis is the biological process of gradual transformation of germ cells into spermatozoa²². Forced swimming stress rats showed a significant decrease in the sperm count, sperm motility and sperm viability which may be due to oxidative stress resulting in decreased spermatogenesis. Low level of cholesterol and testosterone also inhibits spermatogenesis. Since testosterone is required for the growth and development of male reproductive organs, in association with FSH, acts on seminiferous tubules to initiate and maintain spermatogenesis²³. Reduced glycogen levels also inhibit the transformation of germ cells into spermatozoa^{24,25}. Sperm motility was reduced may be due to decreased fructose levels²⁶, inhibiting energetic transferase or non protein substances in the epididymis²⁷.

Abnormal forms of sperms (Fig 1) were seen in forced swimming rats which may be due to oxidative stress, non availability of hormones or any other substances required for maintaining the shape and structure²⁸. Similar results were seen from previous studies, where intensive swimming causes reduction in sperm count and motility²⁹. EESC rhizomes at both doses showed normal sperm count, sperm motility, sperm viability and less or no changes in the sperm morphology may be due to free radical scavenging properties promoting spermatogenesis.

From the experimental studies carried out, ethanolic extract of *Smilax china* rhizomes at two different doses 100 and 200 mg/kg b.w showed free radical scavenging activity and restoration to normal spermatological parameters. These activities may be due to the presence of various phytochemical constituents like saponins, flavonoids, tannins, terpenes, alkaloids and other unknown chemical constituents present in the ethanolic extract of *Smilax china* rhizomes.

Further studies are required to be carried out in isolating the potential chemical constituents present in ethanolic extract of *Smilax china* rhizomes and to find its mechanism of action in treating male reproductive dysfunctions.

REFERENCES

1. Sikka SC, Rajasekaran M, Hellstrom WJ. Role of oxidative stress and antioxidants in male infertility. *J Androl* 1995; 16:464-8.
2. Joyce DA. Oxygen radicals in disease. *Adverse Drug Reaction Bull* 1987; 127:476-9.
3. Tremellen K. Oxidative stress and male infertility-a clinical perspective. *Hum Reprod Update* 2008; 14:243-58.
4. Agarwal A, Makker K, Sharma R. Clinical relevance of oxidative stress in male factor infertility: an update. *Am J Reprod Immunol* 2008; 59:2-11.
5. Terry TT and Jeffrey JL. Oxidative stress: A common factor in testicular dysfunction. *J Androl*. 2008; 29 Suppl 5:488-98.
6. Khare CP. *Indian Medicinal Plants*. New York (NY): Springer; 2007.
7. Chopra RN, Nayar SL, Chopra IC. *Glossary of Indian medicinal plants. Publication and information Directorate, CSIR, New Delhi*; 1956.
8. Xiao SS, Zhong HZ, Xiang LY. Antiinflammatory and antinociceptive activities of *Smilax china* Linn. *J Ethnopharmacol* 2006; 103Suppl 3:327-332.
9. Li YL, Gan GP, Zhang HZ, Wu HZ, Li CL, Huang YP, et al. A flavonoid glycoside isolated from *Smilax china* L. rhizome *in vitro* anticancer effects on human cancer cell lines. *J Ethnopharmacol*. 2007; 113 Suppl 15:115-24.
10. Jieyun J, Qiang X. Immunomodulatory activity of the aqueous extract from the rhizome of *Smilax glabra* in the phase of adjuvant-induced arthritis in rats. *J Ethnopharmacol* 2003; 85 Suppl 1:53-9.
11. Liu Shi-wang, you bi-gang, Yan-xia. Inhibition effect of the ethanol extract of *Smilax China* L. on micro-organisms. *Resource development and market* 2004; 113 Suppl 1:115-2.
12. Health effects test guidelines. Acute oral toxicity [computer program] OPPTS 870.1100 united states of prevention, pesticides and toxic substance environmental protection agency (7101).
13. Debanka SM, Rajkumar M, Saradindu B, et al. Protective extract of *Withania somnifera*, *Ocimum sanctum* and *Zingiber officinale* on swimming-induced reproductive endocrine Dysfunctions in male rat. *Iranian J of Pharmacol and Therapeutics* 2005; 4 Suppl 2:110-7.
14. Ghasem S, Fakher R, Karim A. Effect of forced swimming stress on count, motility and fertilization capacity of the sperm in adult rats. *J of Hum Reprod Sci* 2009; 2 Suppl 2:72-5.
15. Revathi P, Vani B, Sarathchandran I, Kadalmani B, Prakash Shyam K, Palnival K. Reproductive toxicity of *Capparis aphylla* in male albino rats. *Int J Pharm Biomed Res*. 2010; 1Suppl 3:102-12.
16. World Health Organization (WHO) laboratory manual for the examination of human semen and sperm-cervical mucus interaction. 1999, 4th ed. Cambridge university press.
17. Vijayabaskaran M, Yuvaraja KR, Babu G, Sivakumar P, Perumal P, Jayakar B. Hepatoprotective and Antioxidant activity of *Symplocos racemosa* bark extract on DMBA induced Hepatocellular carcinoma in rats. *Inter J Curr Trends Sci Tech* 2010; 1Suppl 3:147-58.
18. Dillard CJ, Litov RE, Tappel AL. Effects of dietary vitamin-E, selenium and polyunsaturated fats on *in vivo* lipid peroxidation in the rats as measured by pentane production. *Indian J Physiol Pharmacol* 1978; 13:396-402.
19. Transler JM, Hales BF, Robaire B. Chronic low dose cyclophosphamide treatment of adult male rats. *Biol Reprod* 1986; 34:275-83.
20. Rai J, Pandey SN, Srivastava RK. Testosterone hormone level in albino rats following restraint stress of long duration. *J Anat Soc India* 2004; 53Suppl 1; 17-9.
21. Peake NJ, Suzuki K, Coombes JS. The influence of antioxidant supplementation on markers of inflammation and the relationship to oxidative stress after exercise. *J Nutr Biochem* 2007; 18:357-71.
22. Sharpe R. Regulation of spermatogenesis. In: Knobil E, Nehil JD, Editors. *The physiology of reproduction*. New York: Raven Press; 1994.p. 1363-434.
23. Mooradan AD, Morely JE, Koreman SG. Biological actions of androgens. *Endo Rev*. 1987;8(1):1-28.
24. Dixit VP, Joshi S. Effects of chronic administration of garlic on testicular function. *Indian J Exp Biol* 1982; 20:534-6.
25. Kusal KD, Shakuntal. Effect of nickel sulfate on testicular steroidogenesis in rats during protein restriction. *Environmental health perspectives*. 2002; 110 Suppl 9: 923-6.
26. Chinoy NJ, Battacharya S. Effects of chronic administration of garlic (*Allium sativum*) on testicular function. *Ind J Exp Biol* 1982; 20:534-6.
27. Pang XB, Zhu Y, Lih G. Effect of ornidazole on sperm in rats and its mechanism of action. *Zhonghua Nan. Ke. Xue* 2005; 11:26-8.

28. D' souza UJ. Effect of tamoxifen on spermatogenesis and tubular morphology in rats. Asian J Androl 2004; 6:223-6.
29. Manna I, Jana K, Samanta PK. Effect of different intensities of swimming exercise on testicular oxidative stress and reproductive dysfunctions in mature male albino Wistar rats. Indian J Exp Biol 2004; 42:816-822.

Source of support: Nil, Conflict of interest: None Declared