



## DNA DAMAGE PROTECTION AND HAEMOLYTIC ACTIVITY OF ISOLONGIFOLENE

Kowsalya R, Jagatheesh. K, Pavan Kumar Padarathi, Elangovan Namasivayam\*

Department of Biotechnology, Periyar University, Salem, Tamil Nadu, India

\*Corresponding Author Email: elangovannn@gmail.com

Article Received on: 29/08/13 Revised on: 22/09/13 Approved for publication: 13/10/13

DOI: 10.7897/2230-8407.041017

IRJP is an official publication of Moksha Publishing House. Website: www.mokshaph.com

© All rights reserved.

## ABSTRACT

In this study a Sesquiterpene, Isolongifolene of *Murraya koenigii* tested for its DNA protecting, genotoxicity and haemolytic activity. DNA Protecting activity was tested against genomic DNA using various concentrations of isolongifolene and the band intensity of each concentration was calculated. Genotoxicity was studied using pUC19 with different concentrations of isolongifolene and the activity was analysed using gel electrophoresis. Haemolytic activity was assayed using human erythrocytes and the activity was measured at 247 nm. Result shows that isolongifolene may protect the genomic DNA from radical damage and it does not produce any toxicity to the pUC19. The haemolytic activity isolongifolene shows the IC<sub>50</sub> value of 50.42 which does not cause haemolysis. Hence, based on the results we conclude that isolongifolene has good protecting activity which can be used for further pharmaceutical studies.

**Keywords:** Isolongifolene; *Murraya koenigii*; DNA damage; genotoxicity; haemolytic activity

## INTRODUCTION

In the normal cells, free radicals and reactive oxygen species (ROS) can be effectively eliminated by an enzyme-mediated system such as superoxide dismutase, peroxidase, glutathione peroxidase and non-enzymatic factors such as ascorbic acid, protein. When the homeostasis between the pro oxidant formation and antioxidant capacity is disrupted, whereby pro oxidant formation exceeds antioxidant capacity, oxidative damage will accumulate and result in patho-physiological events. Accumulating evidence indicates that active oxygen and free radicals would attack key biological molecules such as DNA, protein, and lipid that leading to many degenerative disease conditions<sup>1</sup>. The use of traditional medicine is widespread and plants still present a large source of natural antioxidants that might serve as leads for the development of novel drugs. Several anti inflammatory, anti necrotic, neuro protective, and hepatoprotective drugs have recently been shown to have an antioxidant and/or antiradical scavenging mechanism as part of their activity<sup>2,3</sup>. *Murraya koenigii* (Linn) Spreng is a tropical tree of the family Rutaceae, which is native to India. The leaves used as herb in Ayurvedic medicine. Their properties include much value as an anti-diabetic<sup>4-7</sup>, antioxidant, antimicrobial<sup>8-11</sup>, anti inflammatory<sup>12</sup>, hepatoprotective<sup>13</sup>, anti hypercholesterolemic<sup>14,15</sup>; as well as efficient against colon carcinogenesis. So far, thirteen compounds were identified in *Murraya Koenigii* leaves. Isolongifolene present in *Murraya Koenigii* leaves and it has anti-proliferative activity. The aim of this work was to study the DNA protecting and Haemolytic activity.

## MATERIALS AND METHODS

Isolongifolene, pUC19, H<sub>2</sub>O<sub>2</sub>, tritonX, agarose were purchased from sigma Aldrich.

## DNA Nicking Assay

DNA nicking assay was performed using genomic DNA by the method of lee *et al.*, 2002<sup>16</sup>. Different concentrations ranging from 2-6 mg of isolongifolene was taken in a microfuge tube. To this 10 µl of fenton's reagent (30 mM H<sub>2</sub>O<sub>2</sub>, 50 µM Acetic acid and 80 µM FeCl<sub>3</sub>) was added and incubated for 30 minutes at 37°C. The DNA was analyzed on 0.8 % agarose gel and it was viewed under UV illuminator.

## Genotoxicity assay

Plasmid nicking assay was carried out by the method described by kits *et al.*, 2000<sup>17</sup>. A mixture of 10 µl (2 mg) of isolongifolene (sigma) and pUC19 (0.5 µg) was taken in a microfuge tube and incubated for 2 h pUC19 (0.5 µg) with 215 µM of H<sub>2</sub>O<sub>2</sub> were served as negative control. 10 µl of the tracking dye was added to the tube (40 mM EDTA, 0.05 % bromophenol blue and 50 % (V/V) glycerol). After incubation the mixture was subjected to electrophoresis. The reaction mixture was run on 0.8 % agarose gel for 40 minutes. Ethidium bromide stained gel was viewed and photographed with UV illuminator. Finally the densitometry calculation was made with Gel Quant. NET software.

## Haemolytic assay

Haemolytic activity was evaluated as described previously by Andra *et al.*, 2008<sup>18</sup>. Human erythrocytes washed with Phosphate Buffer Saline (pH 7.4) and centrifuged at 8000 g for 10 minutes. After washing with PBS (until the supernatant was colourless), then the erythrocytes were re-suspended and diluted to 10 times of the original volume with PBS kept as stock. Then, 150 µl of isolongifolene (2-16 µg/ml) in PBS was incubated with 150 µl of stock erythrocyte suspension (4 % v/v) for 60 minutes at 37°C. After the incubation period, the reaction mixtures were centrifuged at 1,000 x g for 10 minutes to remove intact erythrocytes. The 10 fold dilution of the supernatant of released haemoglobin was measured at 540 nm using a micro plate reader. The triplicates experiment was done. Finally the haemolytic activity was expressed as a percentage haemolysis using the following equation. Finally the IC<sub>50</sub> Value was calculated by using PRISM version 5 software.

$$\% \text{ of Haemolysis} = \frac{(\text{Abs sample} - \text{Abs buffer}) / (\text{Abs maximum} - \text{Abs buffer}) \times 100}{100}$$

## RESULTS AND DISCUSSION

Hydroxyl radical is one of the ROS formed in biological systems, causing DNA strand breakage, which leads carcinogenesis, mutagenesis and cytotoxicity<sup>19</sup>.

DNA protective activity of isolongifolene was assessed on hydroxyl radical generated DNA stand breakage was

evaluated using genomic DNA. Different concentration of isolongifolene was treated with fenton's reagent and the protecting activity was shown Figure 1. Band intensity was calculated to analyze the effectiveness of isolongifolene and the result shown in graph 1. The results proved that isolongifolene protects DNA from damage caused by hydroxyl radicals and band intensity revealed that isolongifolene exhibits DNA protecting activity. Genotoxicity of isolongifolene was tested by the method described by kits *et al.*, 2000. The genotoxicity effect of isolongifolene was evaluated with pUC19 plasmid DNA. The genotoxicity effect of isolongifolene and band intensity was shown Figure 2 and graph 2. The plasmid DNA was mainly of super coiled, open circular and linear DNA and isolongifolene results that it does not affect any form of the plasmid DNA. Some of the Plant derived compounds are toxic to the cell. Sesquiterpene lactones are used to treat some diseases which may lyse heme. Isolongifolene was tested for its haemolytic activity. Haemolytic activity of isolongifolene was shown in graph 3. When compared to other plant derived compound saponins the IC<sub>50</sub> value of isolongifolene was found to be 50.42. The results proved that isolongifolene possess insignificant to haemolysis.

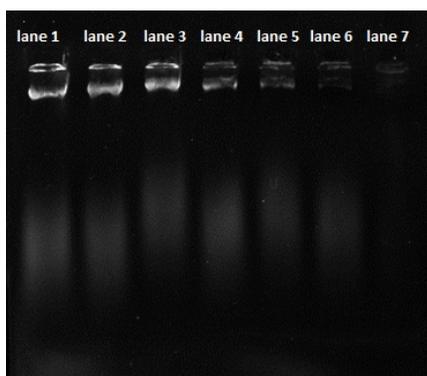
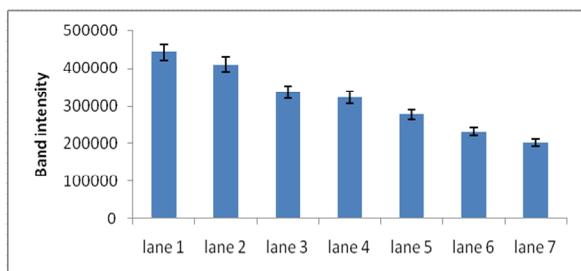


Figure 1: DNA protecting activity of Isolongifolene

Lane 1 - control Genomic DNA  
 Lane 2 - Fenton's reagent + Genomic DNA + quercetin  
 Lane 3 - Fenton's reagent + Genomic DNA + Isolongifolene (8 mg);  
 Lane 4 - Fenton's reagent + genomic DNA + Isolongifolene (6 mg)  
 Lane 5 - Fenton's reagent + Genomic DNA + Isolongifolene (4 mg)  
 Lane 6 - Fenton's reagent + Genomic DNA + Isolongifolene (2 mg)  
 Lane 7- Fenton's reagent + Genomic DNA

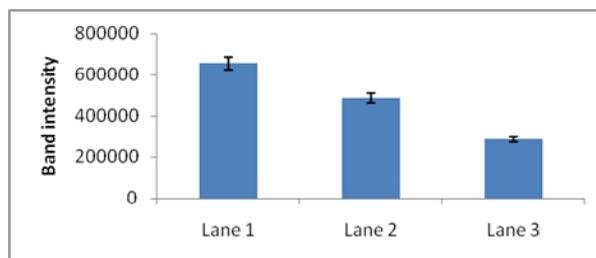


Graph 1: Band Intensity graph of DNA protecting activity

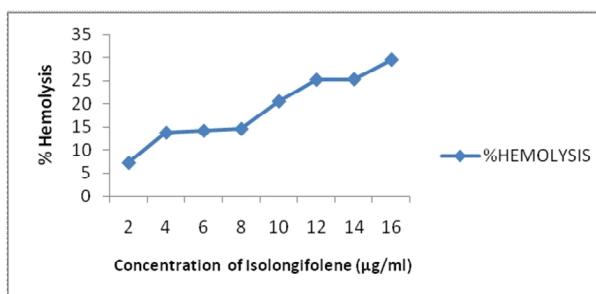


Figure 2: Genotoxicity activity of isolongifolene

Lane 1: control pUC19 plasmid DNA  
 Lane 2: pUC19 Plasmid DNA + 2 mg of isolongifolene  
 Lane 3: pUC19 Plasmid DNA + H<sub>2</sub>O<sub>2</sub>



Graph 2: Band Intensity graph of genotoxicity



Graph 3: Haemolytic activity of Isolongifolene

## CONCLUSION

Isolongifolene does not cause damage, toxicity to DNA and it may not lyse the heme. The results conclude that isolongifolene has good protecting activity. Hence isolongifolene is safe for use and it can be further studied for its activity.

## REFERENCES

- Suja KP, Jayalekshmy A and Arumughan C. Free radical scavenging behaviour of antioxidant compounds of sesame (*Sesamum indicum* L.) in DPPH system. *J. Agric. Food Chem* 2004; 52: 912-915. <http://dx.doi.org/10.1021/jf0303621> PMID:14969550
- Perry EK, Pickering AT, Wang WW, Houghton PJ and Perru NS. Medicinal plants and Alzheimer's disease: from ethno botany to phytotherapy. *J. Pharm. Pharmacology* 1999; 51: 527-534. <http://dx.doi.org/10.1211/0022357991772808> PMID:10411211
- Repetto MG, Llesuy SF. Antioxidant properties of natural compounds used in popular medicine for gastric ulcers. *Braz J. Med. Biol. Res* 2002; 35: 523-534. <http://dx.doi.org/10.1590/S0100-879X2002000500003> PMID:12011936
- Arulselvan P, Senthilkumar GP, Sathish Kumar D and Subramanian S. Anti-diabetic effect of *Murraya koenigii* leaves on streptozotocin induced diabetic rats. *Pharmazie* 2006; 61(10): 874-877. PMID:17069429
- Yadav S, Vats V, Dhunnoo Y and Grover JK. Hypoglycemic and anti hyperglycemic activity of *Murraya koenigii* leaves in diabetic rats. *J Ethnopharmacol* 2001; 82: 111. [http://dx.doi.org/10.1016/S0378-8741\(02\)00167-8](http://dx.doi.org/10.1016/S0378-8741(02)00167-8)
- Vinuthan MK, Girish Kumar V, Ravindra JP and Narayana K. Effect of extracts of *Murraya koenigii* leaves on the levels of blood glucose and

- plasma insulin in alloxan induced diabetic rats. Indian J Physiol Pharmacol 2004; 48: 348. PMID:15648408
7. Achyut NK, Gupta RK and Watal G. Hypoglycemic effects of *Murraya koenigii* on normal and alloxan-diabetic rabbits. J. Ethno pharmacology 2005; 97: 247. <http://dx.doi.org/10.1016/j.jep.2004.11.006> PMID:15707761
  8. Arulselvan P and Subramanian SP. Beneficial effects of *Murraya koenigii* leaves on antioxidant defence system and ultra structural changes of pancreatic beta-cells in experimental diabetes in rats. Chem Biol Interact 2007; 165(2): 155–64. <http://dx.doi.org/10.1016/j.cbi.2006.10.014> PMID:17188670
  9. Singh L and Sharma M. Antifungal properties of some plant extracts. Geobios 1978; 5: 49.
  10. Baliga MS, Jagetia GC, Rao SK and Babu K. Evaluation of nitric oxide scavenging activity of certain spices *in vitro*: a preliminary study. Nahrung 2003; 47(4): 261. <http://dx.doi.org/10.1002/food.200390061> PMID:13678266
  11. Mathur Abhishek, Dua VK and Prasad GBKS. Antimicrobial Activity of Leaf Extracts of *Murraya Koenigii* against Aerobic Bacteria Associated with Bovine Mastitis. Int. J. of Chemical Environmental and Pharmaceutical Research 2010; 1(1): 12-16.
  12. Muthumani P, Venkatraman S, Ramseshu KV, Meera R, Devi P, Kameswari B and Eswarapriya B. Pharmacological studies of anticancer, anti inflammatory activities of *Murraya Koenigii* (Linn) Spreng in experimental animals. J. Pharm. Sci. and Re 2009; 1(3): 137-141.
  13. Arulmozhi S, Papiya A, Purnima A and Sathiya N. *In vitro* antioxidant and free radical scavenging activity of *Alstonia scholaris* Linn. Iranian Journal of Pharmacology and Therapeutics 2008; 6: 191-196.
  14. Pande M, Gupta SPBN and Pathak A. Hepatoprotective activity of *Murraya Koenigii* Linn. Bark. Journal of Herbal Medicine and Toxicology 2009; 3(1): 69-71.
  15. Iyer UM and Mani UV. Studies on the effect of curry leaves supplementation (*Murraya koenigii*) on lipid profile, glyated proteins and amino acids in non-insulin dependent diabetic patients. Plants and Food in Human Nutrition 199; (40): 275.
  16. Lee JC, Kim HR, Kim J and Jang YS. Antioxidant property of an ethanol extract of the stem of *Opuntia ficus-indica* var. Saboten. Journal of Agricultural and Food Chemistry 2002; 50(22): 6490–6496. <http://dx.doi.org/10.1021/jf020388c> PMID:12381138
  17. Kitts CL, Green CE, Otley RA, Alvarez MA, and Unkefer PJ. Type I nitro reductases in soil entero bacteria reduce TNT (2, 4, 6,-trinitrotoluene) and RDX (hexahydro-1, 3, 5-trinitro-1, 3, 5-triazine). Can J Microbiol 2000; 46(3): 278-82. <http://dx.doi.org/10.1139/w99-134> PMID:10749541
  18. Andra J, Jakovkin I, Grotzinger J, Hecht O, Krasnosdembskaya AD and Goldmann T. Structure and mode of action of the antimicrobial peptide arenicin. Biochem J 2008; 410:113-22. <http://dx.doi.org/10.1042/BJ20071051> PMID:17935487
  19. Prieto P, Pineda M and Aguilar M. Spectrophotometric quantitation of antioxidant capacity through the formation of phosphomolybdenum complex: specific application to determination of vitamin E. Analytical Biochemistry 1999; 269: 337–341. <http://dx.doi.org/10.1006/abio.1999.4019> PMID:10222007
- Cite this article as:**  
Kowsalya R, Jagatheesh. K, Pavan Kumar Padarthi, Elangovan Namasivayam. DNA damage protection and haemolytic activity of Isolongifolene. Int. Res. J. Pharm. 2013; 4(10):75-77 <http://dx.doi.org/10.7897/2230-8407.041017>

Source of support: Nil, Conflict of interest: None Declared